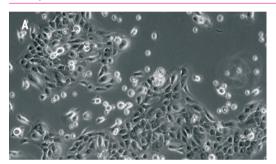


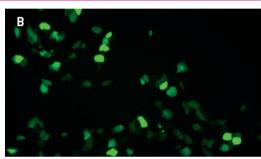
Amaxa™ Nucleofector™ Protocol for Human Prostate Epithelial Cells (PrEC)

For Human Prostate Epithelial Cells (hPrEC)

Validated to work with Clonetics™ PrEC [Lonza; Cat. No. CC-2555]; adherent epithelial cells

Example for Nucleofection™ of human PrECs





Human PrECs were transfected using program W-001 and a plasmid encoding the green fluorescent protein, maxGFP® Vector. 24 hours post Nucleofection™ the cells were analyzed by light (A) or fluorescence microscopy (B).



Transfection efficiency of hPrECs 24 hours post Nucleofection. Cells were transfected using program W-001 and 2 μ g of pmaxGFP $^{\infty}$ Vector..

Product Description

Recommended Kit(s): Basic Nucleofector™ Kit for Primary Mammalian Epithelial Cells

Cat. No.	VPI-1005	
Size (reactions)	25	
Nucleofector™ Solution	2.25 ml (2.05 ml + 10% overfill)	
Supplement	0.5 ml (0.45 ml + 10% overfill)	
pmaxGFP™ Vector (0.5 μg/μl in 10 mM Tris pH 8.0)	30 μg	
Certified cuvettes	25	
Plastic pipettes	25	

Storage and stability

Store Nucleofector™ Solution, Supplement and pmaxGFP™ Vector at 4°C. For long-term storage, pmaxGFP™ Vector is ideally stored at -20°C. The expiration date is printed on the solution box. Once the Nucleofector™ Supplement is added to the Nucleofector™ Solution it is stable for three months at 4°C.

Optimized Protocol for Human Prostate Epithelial Cells

Required Material

Note

Please make sure that the entire supplement is added to the Nucleofector™ Solution. The ratio of Nucleofector™ Solution to supplement is 4.5: 1. For a single reaction use 82 µl of Nucleofector™ Solution plus 18 µl of supplement to make 100 µl of total reaction volume.

- Nucleofector™ Device
- Supplemented Nucleofector™ Solution at room temperature
- Supplied certified cuvettes
- Supplied plastic pipettes
- Supplied pmaxGFP™ Vector
- Substrate of interest, highly purified, preferably by using endotoxin free kits; A260: A280 ratio should be at least 1.8
- For trypsinization: Reagent Pack™ Subculture Reagent Kit containing Trypsin/EDTA, HEPES Buffered
 Saline Solution (HBSS) and Trypsin Neutralizing Solution (TNS) [Lonza; Cat. No. CC-5034]
- Prepared 6-well culture dish
- Culture medium: PrEGM® BulletKit [Lonza; Cat. No. CC-3166] per reaction). We recommend storing 40 ml aliquots of the prepared medium at -80°C. Do not use medium stored at 4°C for more than two days, as this may lead to increased cell mortality and significant reduction of transfection efficiency
- Prewarm appropriate volume of culture media at 37°C (1.5 ml per reaction)
- Appropriate number of cells (5 x 10⁵ cells per sample); minimal cell number: 3 x 10⁵ cells (a lower cell number may lead to a major increase in cell mortality); maximal cell number: 7 x 10⁵ cells

1. Pre Nucleofection™

Note

Transfection results may be donor-dependent.

Cell culture recommendations

- 1.1 Seeding conditions: 2.5 x 10³ cells/cm². Use flasks with a surface area of 75 m² only. High cell densities in hPrEC culture lead to reduced cell viability and transfection efficiency
- 1.2 Replace medium 1 day after splitting, then every 2 days
- 1.3 Cells should be passaged every 3 4 days
- 1.4 For Nucleofection™ cells should be preferably passaged 2 days before
- 1.5 Do not use cells after passage number 8 as this may result in substantially lower gene transfer efficiency and viability
- 1.6 Optimal confluency before Nucleofection™ 60 80%

Trypsinization

- 1.7 Remove media from the cultured cells and wash cells once with HBSS; use at least same volume of HBSS as culture media
- 1.8 For harvesting, incubate the cells 4 6 minutes at 37°C with recommended volume of indicated trypsinization reagent (please see required material)
- 1.9 Neutralize trypsinization reaction with TNS once the majority of the cells (>90%) have been detached (latest after 7 minutes as otherwise cells may start to clump)

Optimized Protocol for Human Prostate Epithelial Cells

2. Nucleofection™

Note

When using self-isolated PrECs or cells obtained from another supplier than Lonza we recommend testing the programs of our basic protocol for primary mammalian epithelial cells: S-005, T-013, T-020, W-001, U-017

One Nucleofection™ Sample contains

5 x 105 cells

 $1-5 \mu g$ plasmid DNA (in $1-5 \mu l$ H₂0 or TE) or $2 \mu g$ pmaxGFP^m Vector or 30-300 nM siRNA $(3-30 \mu l)$ pmol/sample)

100 µl Nucleofector™ Solution

- 2.1 Please make sure that the entire supplement is added to the Nucleofector™ Solution
- 2.2 Prepare 6-well plates by filling appropriate number of wells with 1.5 ml of supplemented culture media and pre-incubate/equilibrate plates in a humidified $37^{\circ}\text{C}/5\%$ CO₂ incubator
- 2.3 Harvest the cells by trypsinization (please see 1.7 1.9)
- 2.4 Count an aliquot of the trypsinized cells and determine cell density
- 2.5 Centrifuge the required number of cells (5 x 10^5 cells per sample) at 220xg for 5 minutes at room temperature
- 2.6 Resuspend the cell pellet carefully in 100 µl room temperature Nucleofector™ Solution per sample
- 2.7 Combine 100 μ l of cell suspension with 1 5 μ g DNA, 2 μ g pmaxGFP^m Vector or 30 nM 300 nM siRNA (3 30 pmol/sample) or other substrates
- 2.8 Transfer cell/DNA suspension into certified cuvette (sample must cover the bottom of the cuvette without air bubbles). Close the cuvette with the cap
- 2.9 Select the appropriate Nucleofector™ Program W-001 (W-01 for Nucleofector™ I Device)
- 2.10 Insert the cuvette with cell/DNA suspension into the Nucleofector™ Cuvette Holder and apply the selected program
- 2.11 Take the cuvette out of the holder once the program is finished
- 2.12 Add \sim 500 μ l of the pre-equilibrated culture media to the cuvette and **gently** transfer the sample into the 6-well plate. Use the supplied pipettes and avoid repeated aspiration of the sample

3. Post Nucleofection™

3.1 Incubate the cells in a humidified 37°C/5% $\rm CO_2$ incubator until analysis. Gene expression is often detectable after only 4 - 8 hours

Optimized Protocol for Human Prostate Epithelial Cells

Additional Information

For an up-to-date list of all Nucleofector™ References, please refer to: www.lonza.com/nucleofection-citations

For more technical assistance, contact our Scientific Support Team:

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