

Instructions for Culturing Gamma Delta ($\gamma\delta$) T Cells Using TheraPEAK® T-VIVO® Cell Culture Medium

I. Introduction

TheraPEAK® T-VIVO® Cell Culture Medium

TheraPEAK® T-VIVO® Cell Culture Medium is optimized to support cell therapy applications utilizing human T cells. It only uses recombinant human proteins and does not require serum or serum substitute. TheraPEAK® T-VIVO® Cell Culture Medium is manufactured in compliance with cGMP and all ingredients are chemically-defined and non-animal origin (NAO). It does not contain cytokines, antibiotics or phenol red.

Gamma delta ($\gamma\delta$) T cells

Gamma delta ($\gamma\delta$) T cells represent a relatively small subset of T cells and are defined by the expression of heterodimeric T-cell receptors (TCRs) composed of γ and δ chains, and can be further identified by different V δ (variable δ) chains. For example, V δ 1 T cells are predominant in the peripheral tissues and V δ 2 T cells constitute the majority of blood $\gamma\delta$ T cells.

$\gamma\delta$ T cells have attractive properties for cancer immunotherapy including their inherent ability to infiltrate tissues and to directly recognize and kill transformed cells independent of HLA-antigen presentation. Moreover, $\gamma\delta$ T cells provide a platform for allogeneic cell therapies as they do not cause graft-versus-host disease (GVHD).

The first protocol describes a process that uses zoledronic acid as the activating agent and peripheral blood mononuclear cells (PBMCs) as the starting cell source for V δ 2 T cells expansion, using TheraPEAK® T-VIVO® Medium without the addition of human serum. V δ 2 T cells can be expanded by more than 1,000-fold in 14 days using this protocol.

A second protocol describes an isolation process that is tailored to deplete $\alpha\beta$ T cells from PBMCs and then uses anti-CD3/anti-CD28 co-stimulation to activate and expand $\gamma\delta$ T cells (V δ 1 and V δ 2 T-cell subsets) using the TheraPEAK® T-VIVO® Medium.

II. Storage

TheraPEAK® T-VIVO® Cell Culture Medium should be stored at 2 to 8 °C, protected from light.

III. Instructions for use

Media preparation

TheraPEAK® T-VIVO® Cell Culture Medium should be supplemented with cytokines, such as IL-2, to support $\gamma\delta$ T-cell expansion. The amount of cytokines required may vary depending on the user applications, however for this protocol, it is suggested to use 100 IU/mL of recombinant human IL-2. The medium with cytokine may be stored at 2 to 8 °C for up to 7 days. When in use, minimize exposure of medium to light.

General guideline for V δ 2 T-cells activated with zoledronic acid and cultured in T-flasks

For optimal gas exchange in static T-flasks, it is recommended that the medium height be less than 3mm.

1. Prepare fresh peripheral blood mononuclear cells (PBMCs) or thaw frozen PBMCs in a 37 °C water bath according to standard thawing protocols.

2. Wash the cells with TheraPEAK® T-VIVO® Cell Culture Medium.
 3. Centrifuge the cells at 200 – 300 x g for 5–10 minutes and remove wash buffer.
 4. Resuspend the cells in TheraPEAK® T-VIVO® Cell Culture Medium with cytokine. Determine viable cell concentration and cell viability using standard cell counting protocols.
 5. Plate 2.0 x 10e6 viable PBMCs in 1 mL TheraPEAK® T-VIVO® Cell Culture Medium with cytokine into one well of a 24-well plate for small scale expansion. For larger cell numbers, plate at 2.0 x 10e6 viable PBMCs/mL of medium in the appropriate vessel to allow for sufficient gas exchange.
 6. Stimulate Vδ2 T cells by adding zoledronic acid to final concentration of 5 – 10 μM.
 7. Incubate the culture vessel at 37 °C in a humidified incubator with 5% CO₂.
Every 2 – 3 days, change 50% of the medium.
 8. When the cells become confluent, transfer the entire culture into a larger vessel and add fresh medium to the appropriate volume.
 9. Continue to expand the T-cell culture by adding fresh TheraPEAK® T-VIVO® Cell Culture Medium with cytokine every 2 – 3 days and re-adjusting the cell density to 1.0 x 10e6 viable cells/mL. Use larger T-flasks as needed.
 10. Harvest cells when the desired cell number is achieved and proceed to downstream applications.
2. Wash the cells with TheraPEAK® T-VIVO® Cell Culture Medium.
 3. Centrifuge the cells at 200 – 300 x g for 5 – 10 minutes and remove wash buffer.
 4. Resuspend the cells in TheraPEAK® T-VIVO® Cell Culture Medium. Determine viable cell concentration and cell viability using standard cell counting protocols.
 5. If start with cryopreserved PBMCs, allow PBMCs recover in TheraPEAK® T-VIVO® Cell Culture Medium overnight at 37 °C in a humidified incubator with 5% CO₂, before isolating the γδ T cells.
 6. Isolate γδ T cells from PBMCs.
A variety of commercial cell separation products may be used to isolate γδ T cells from PBMCs. Non-γδ T cells, i.e. αβ T cells, NK cells, B cells, dendritic cells, granulocytes, monocytes, stem cells etc. may be depleted using a cocktail of antibodies. At minimum, αβ T cells should be depleted using αβTCR-specific antibodies.
 7. Determine viable cell concentration of the isolated γδ T cells using standard cell counting protocols.
 8. For research purposes, plate 0.5 – 1.0 x 10e6 viable γδ T cells in 1 mL TheraPEAK® T-VIVO® Cell Culture Medium with cytokine into one well of a 24-well plate. Alternatively, a 96-well plate may be used, in which case, plate 5.0 – 10.0 x 10e3 viable cells in one well of a 96-well plate in 150 μL medium.
If only αβ T cells are depleted from PBMCs, plate 1.0 – 2.0 x 10e6 viable cells in 1 mL TheraPEAK® T-VIVO® Cell Culture Medium with cytokine into one well of a 24-well plate for small scale expansion.
 9. Stimulate T cells for expansion using commercial anti-CD3 and anti-CD28 T cell activation products as recommended by the suppliers. For example, use 10μL T Cell TransAct™ (Miltenyi) for 1 mL medium.
 10. Incubate the culture vessel at 37 °C in a humidified incubator with 5% CO₂.
 11. Continue to expand the T-cell culture by adding fresh TheraPEAK® T-VIVO® Cell Culture Medium with cytokine every 2 – 3 days.

General guideline for γδ T-cell expansion with anti-CD3/anti-CD28 co-stimulation after depleting αβ T cells from PBMCs

Both γδ T cells and αβ T cells express CD3 and CD28, hence it is critical to deplete αβ T cells prior to expansion using this protocol. αβ T cells may be removed from PBMCs with a variety of commercial cell separation products.

1. Prepare fresh peripheral blood mononuclear cells (PBMCs) or thaw frozen PBMCs in a 37 °C water bath according to standard thawing protocols.

Count the cells if possible and re-adjust the cell density to 0.5 – 1.0 x 10⁶ viable cells/mL. Use larger T-flasks as needed.

12. Harvest cells when the desired cell number is achieved and proceed to downstream applications.

IV. General recommendations

Refer to the general TheraPEAK® T-VIVO® Cell Culture Medium instruction for recommendations on adopting processes that maintain good dissolved oxygen levels during the cell expansion processes, and optimizing processes that involve Nucleofection® or electroporation.

Lentivirus transduction in $\gamma\delta$ T cells

Lentivirus pseudotyped with the common VSV-G envelope protein enters cells using the low-density lipoprotein receptor (LDL-R) as the major entry receptor, however, LDL-R expression is very low in resting T cells. Stimulation of T cells through the T-cell receptor upregulates the LDL-R expression within 24-72 hours after stimulation, and permits efficient VSV-G pseudotyped lentivirus transduction. Therefore, for optimal lentivirus transduction of $\gamma\delta$ T cells, it is recommended to activate the cells by crosslinking CD3 and CD28 followed by lentivirus transduction between 24 and 72 hours after activation.

Product use statement

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