

# Lonza



## PyroGene® Recombinant Factor C Endotoxin Detection Assay

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## Use

PyroGene® rFC Assay is to be used only as an *in vitro* end-product endotoxin test for human and animal parenteral drugs, biological products, and medical devices. This product is not intended for use in the detection of endotoxin in clinical samples for the diagnosis of human disease.

The PyroGene® Test utilizes recombinant Factor C (rFC), an endotoxin-sensitive protein. rFC is used in combination with a fluorogenic substrate, an incubating fluorescence microplate reader and appropriate software to measure endotoxin. A minimum detection limit of 0.005 EU/mL and a measurable endotoxin concentration range of 0.005 to 5.0 EU/mL can be achieved.

## Warning

For *In Vitro* Use Only. The PyroGene® Assay is not intended to detect endotoxemia in man or animals, or for use in clinical diagnosis, patient management, or for the qualification of blood or blood products.

## Safety Precautions

The toxicological properties of the reagents supplied have not been tested. These reagents are not considered hazardous according to the OSHA Hazard Communication Standard 29 CFR 1910.1200. It is recommended that the NIH guidelines for recombinant DNA experiments be followed along with the use of standard laboratory precautions.

## Background

A Gram-negative bacterial infection of *Limulus polyphemus*, the horseshoe crab, may result in fatal intravascular coagulation<sup>1</sup>. At the molecular level, it has been demonstrated that endotoxin activates a serine protease catalytic coagulation cascade that results in the gelation of *Limulus* blood. This cascade is used in the *Limulus* Amebocyte Lysate (LAL)<sup>1, 2, 3, 4</sup> endotoxin detection method.

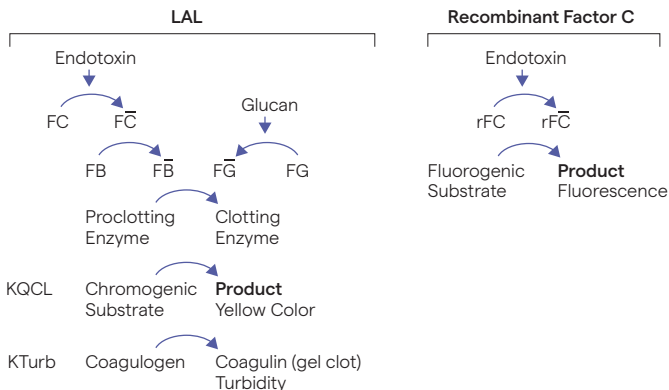
The protease cascade and rationale of traditional LAL tests are illustrated in the LAL Pathway, Figure 1 (next page).

Factor C, the first component in the cascade, is a protease zymogen that is activated by endotoxin binding<sup>5, 6, 7</sup>. In this pathway, Factor B (FB) is activated by Factor C. An alternative pathway, the Factor G (FG) pathway, can be activated by glucan binding<sup>8</sup>. Downstream, Factor C and Factor G pathways individually activate a proclotting enzyme into a clotting enzyme. The chromogenic LAL assay (Lonza's Kinetic-QCL<sup>®</sup> Assay) uses a synthetic chromogenic peptide substrate that can be cleaved by the clotting enzyme, resulting in a product that exhibits a yellow color. The turbidimetric assay (Lonza's PYROGENT<sup>®</sup> 5000 Assay, KTurb) uses the native substrate, coagulogen, which can be cleaved into coagulin. Coagulin then begins to self-associate, resulting in an increase in turbidity. The densities of the yellow color (OD 405 nm) and turbidity (OD 340 nm) are correlated with endotoxin concentration.

Studies have demonstrated the ability of Factor C to selectively recognize endotoxin and activate the protease

cascade. To create an endotoxin-specific assay, Factor C has been purified and cloned<sup>5, 6, 7, 9, 10, 11</sup>. When activated by endotoxin binding, recombinant Factor C acts upon the fluorogenic substrate in the assay mixture to produce a fluorescent signal in proportion to the endotoxin concentration in the sample.

### Endotoxin Detection



**Figure 1.** Schematic drawing of the endotoxin detection mechanisms in the LAL system and the rFC system.

## Principle

Recombinant Factor C is activated by endotoxin binding, and the active moiety created then acts to cleave a synthetic substrate resulting in the generation of a fluorogenic compound (Figure 1, rFC pathway). The assay is carried out in a 96-well plate. Fluorescence is measured at time zero and after a one-hour incubation at  $37^{\circ}\text{C} \pm 1^{\circ}\text{C}$  in a fluorescence microplate reader using excitation/emission wavelengths of 380/440 nm. The difference between the one-hour reading and the time zero reading ( $\Delta\text{RFU}$ ) is corrected for blank  $\Delta\text{RFU}$  fluorescence. The log net fluorescence is proportional to the log endotoxin concentration and is linear in the 0.005 to 5.0 EU/mL range. Endotoxin in a sample is calculated relative to a standard curve.

## Storage Conditions

1. The PyroGene<sup>®</sup> Kit, catalog numbers 50-658U, 50-658NV, is stored at 2 to 8°C.

## Reagents Supplied and Kit Configurations

### Catalog No. 50-658U contains:

1. Two vials of rFC Enzyme Solution, R50-658, 1.2 mL/vial
2. Two vials of the Fluorogenic Substrate, S50-658, 6.0 mL/vial
3. Two vials of the rFC Assay Buffer, B50-658, 5.0 mL/vial
4. Two vials of the *E. coli* Endotoxin, O55:B5, E50-643, lyophilized. The reconstitution volume of the vial is stated on the Certificate of Analysis and is calculated to yield a solution containing 20 EU/mL. Reconstituted endotoxin is stable for 4 weeks at 2 to 8°C.

5. Two vials of LAL Reagent Water, W50-640, 30 mL/vial. This water is used to rehydrate *E. coli* endotoxin and to prepare endotoxin standard and sample dilutions. LAL Reagent Water is equivalent to Water for Bacterial Endotoxins Test (BET).

**Catalog No. 50-658NV contains:**

1. Thirty vials of rFC Enzyme Solution, R50-658, 1.2 mL/vial
2. Thirty vials of the Fluorogenic Substrate, S50-658, 6.0 mL/vial
3. Thirty vials of the rFC Assay Buffer, B50-658, 5.0 mL/vial
4. Ten vials of the *E. coli* Endotoxin, O55:B5, E50-643, lyophilized. The reconstitution volume of the vial is stated on the Certificate of Analysis and is calculated to yield a solution containing 20 EU/mL. Reconstituted endotoxin is stable for 4 weeks at 2 to 8°C.

The COA is available at [www.lonza.com/coa](http://www.lonza.com/coa).



## Materials and Equipment Required but NOT Supplied

1. Disposable endotoxin-free glass dilution tubes (13 × 100 mm, N207 or equivalent).
2. Individually wrapped sterile measuring pipettes.
3. Automatic hand-held pipettes with sterile, individually wrapped or racked tips.
4. Disposable sterile microplates. Note: Prior to routine use, microplates should be pre-qualified (25-340 or equivalent). Black-walled microplates are recommended for use in fluorescence-based assays [Corning 3603 or equivalent]
5. Reagent reservoirs (00190035 or equivalent).
6. Eight channel pipettor.
7. Fluorescence microplate reader (25-375S Nebula<sup>®</sup> Multimode Reader, 25-345S PyroWave<sup>®</sup> XM Fluorescence Reader with Ex 380/20 and Em 440/30, or equivalent).
8. Software (WinKQCL<sup>®</sup> V 5.3.3, or higher, recommended).
9. Timer.
10. Vortex mixer.
11. For 50-658NV, LAL Reagent Water (W50-640 or equivalent). LAL Reagent Water is equivalent to Water for Bacterial Endotoxins Test (BET).

## Specimen Collection and Preparation

Careful technique must be used to avoid microbiological or endotoxin contamination. All materials coming in contact with the specimen or test reagents must be endotoxin-free. Clean glassware and materials may be rendered endotoxin-free by heating at 250°C for 30 minutes. Appropriate precautions should be taken to protect depyrogenated materials from subsequent environmental contamination.

From experience, most sterile, individually wrapped, plastic pipettes and pipette tips are endotoxin-free. However, these materials should be tested before regular use.

It may be necessary to adjust the pH of the sample to within the range 6.0 – 8.0 using endotoxin-free sodium hydroxide or hydrochloric acid. Always measure the pH of an aliquot of the bulk sample to avoid contamination by the pH electrode. Do not adjust unbuffered solutions.

Samples to be tested must be stored in such a way that all bacteriological activity is stopped or the endotoxin level may increase with time. To suppress bacteriological activity, store samples at 2 to 8°C for less than 24 hours and frozen for periods greater than 24 hours. It is the responsibility of the customer to validate the proper container and storage conditions for their samples.

## Sensitivity Setting for Fluorescent Reader

Fluorescence signals are normally recorded as Relative Fluorescence Units (RFU). Since the real fluorescence signal is converted to an electronic signal that can be “tuned” using the gain setting or sensitivity setting, RFU is an arbitrary unit. Depending on the strength of the detected signal, the instrument can be adjusted to a higher gain/sensitivity setting to boost a weak signal or tuned down to a lower gain/sensitivity setting when the signal is too strong. It is recommended to consult the reader User’s Manual for the manufacturer’s explanation.

In the PyroGene® Assay, the 3-log endotoxin concentration range corresponds to the 3-log range in RFU. If the sensitivity is tuned too low, fluorescence at the lowest standard will be difficult to detect; if the sensitivity is tuned too high, fluorescence at the highest standard will be off-scale. Prior to conducting any assays, it is important to determine the appropriate sensitivity for the PyroGene® Assay. For example, the FLx800™ and PyroWave® XM, readers have a fluorescence range from 0 to 99,999. The Nebula® Multimode Reader has a fluorescence range from 0 to 65,535. In order to establish the 3-log fluorescence range for the standard curve, an RFU range from 1,000 to 10,000 was chosen for the 0.5 EU/mL standard. The RFU range from 1,000 to 10,000 corresponds to a log range from 3 to 4, with a log mid-point of 3.5. Thus, the target absolute net RFU for the 0.5 EU/mL is approximately 3,000 RFU. The PyroWave® XM Reader is more reliable over the entire dynamic range which results in the PyroGene® Assay producing a slope of

<1.00. To account for this difference a higher log mid-point target was set so that the reader will solve for a log value of 4.00 when determining the sensitivity setting. The Target delta RFU for the Nebula® MMR is 3.7. To maintain adequate separation between the blanks and lowest standard in a routine assay, this value should be considered the minimum desirable range. Consult the appropriate User's Manual for recommendations for alternative readers.

### **Determination of Appropriate Sensitivity Setting**

Note: This procedure should be conducted as part of the performance qualification for any new lot of reagents, after major equipment maintenance (such as lamp replacement) or when reader performance is questioned. A periodic re-verification is recommended as part of any on-going equipment maintenance program. It is recommended to reassess the sensitivity setting on each reader at least every 3 – 6 months.

1. Prepare a 0.5 EU/mL endotoxin solution (see Preparation of Endotoxin Standards). Allow reagents to equilibrate to ambient temperature before mixing.
2. See WinKQCL® Software Manual or Help for prompts to the Sensitivity Test.
3. Enter appropriate information in blank fields and confirm settings for the test. The example on the following page compares settings for the FLx800™ Reader, the PyroWave® XM Reader, and the Nebula® Multimode Reader. Critical differences in settings for each reader are in purple.

	FLx800™	PyroWave® XM	Nebula® MMR
Endotoxin Concentration Unitage	EU	EU	EU
Time between reads (hh:mm:ss)	01:00:00	01:00:00	01:00:00
Excitation filter (nm)	380:20	380:20	380:9
Emission filter (nm)	440:30	440:30	440:20
Scans per well	4	4	10
Delay before scanning (millisec.)	150	<b>200</b>	<b>200</b>
Delay between scans (millisec.)	1	1	N/A
Optics position	bottom	<b>top</b>	<b>top</b>
Wells to use: Pre-defined	(D6,7; E6,7; F6,7)	(D6,7; E6,7; F6,7)	(D6,7; E6,7; F6,7)
Sensitivity (select by checking)	<b>30, 35, 40, 45, 50, 55</b>	<b>25, 28, 31, 34, 37, 40</b>	<b>57, 62, 67, 72, 77, 82</b>

- Add 100 µL of the 0.5 EU/mL solution to wells D6,7; E6,7; F6,7.
- Pre-incubate covered plate with standards at 37°C ± 1°C for 10 minutes.
- Prepare working reagent by mixing the fluorogenic substrate, rFC assay buffer and the rFC enzyme solution at 5:4:1 ratios, respectively. 500 µL + 400 µL + 100 µL is a sufficient quantity for the sensitivity test. Refrigerate any unused reagents after opening. Previously opened vials remain effective throughout the lifetime of the reconstituted endotoxin if returned to refrigeration. Acceptable assay evaluation parameters will indicate

proper reagent performance. Note: The order of mixing should remain consistent. Add enzyme last to the buffered substrate. Mix thoroughly but gently. Do not vortex the working reagent mix. Note: Once prepared, the working reagent cannot be stored.

7. At the appropriate software prompt, add 100  $\mu\text{L}$  of working reagent to all wells that contain the endotoxin sample. Follow screen prompt to initiate test. Note: Leave the plate uncovered for the duration of the test.
8. At the completion of the test a summary of the results with the calculated sensitivity will be displayed. Using linear regression, the sensitivity value correlated to  $\log \Delta\text{RFU} = 4.0$  ( $\log \Delta\text{RFU} = 3.5$  for the FLx800™ Reader) is rounded to the nearest integer. Successful assays should result when using this setting.

Note: If the predicted sensitivity result is outside of the range of the tested default parameters of 25, 28, 31, 34, 37, 40 (default range for the FLx800™ Reader is 30, 35, 40, 45, 50 and 55), then the sensitivity assay should be repeated utilizing a more appropriate range that will include that result. It is not recommended to extrapolate beyond the range of the curve.

Sensitivity Setting Determined: \_\_\_\_\_

For Kit Lot: \_\_\_\_\_

## Preparation of Endotoxin Standards

In order to calculate the endotoxin concentration in unknown samples, each PyroGene® Test must be referenced to a valid standard curve.

Because of the large concentration range over which endotoxin values can be determined, it is possible to adjust the quantitative range of any given test by adjusting the concentration of endotoxin standards used to generate the standard curve. A minimum of three standards is required.

The PyroGene® Assay has been optimized to be linear from 0.005 EU/mL to 5.0 EU/mL. However, the individual user may choose to truncate the standard curve depending on specific product testing requirements.

The *E. coli* endotoxin, O55:B5, is provided for user convenience. Other endotoxin preparations may be used to prepare the standards; however, their performance in the PyroGene® Assay relative to the current USP Reference Standard Endotoxin (RSE) must be determined.

The supplied lyophilized *E. coli* O55:B5 endotoxin is reconstituted with LAL Reagent Water to yield a 20 EU/mL stock solution. The reconstitution volume of the vial is stated on the Certificate of Analysis and is calculated to yield a solution containing 20 EU/mL. Reconstitute with the specified volume of LAL Reagent Water. Shake vigorously for approximately 15 minutes at high speed on a vortex mixer. Prior to future use, the solution must be warmed to room temperature and vigorously vortexed for approximately 15 minutes.

The following table suggests a dilution scheme for constructing a series of endotoxin dilutions from the endotoxin supplied in the kit.

Note: Plastic tubes are not recommended for making endotoxin dilutions.

Endotoxin Concentration (EU/mL)	Volume of LAL Reagent Water	Standard Added to LAL Reagent Water
5.0	0.75 mL	0.25 mL of 20 EU/mL solution
0.5	0.9 mL	0.1 mL of 5.0 EU/mL solution
0.05	0.9 mL	0.1 mL of 0.5 EU/mL solution
0.005	0.9 mL	0.1 mL of 0.05 EU/mL solution

1. Prepare a solution containing 5.0 EU/mL endotoxin by adding 0.25 mL of the 20 EU/mL endotoxin stock solution into 0.75 mL of LAL Reagent Water in a suitable container and label 5.0 EU/mL. This solution should be vigorously vortexed for approximately 1 minute before proceeding.
2. Transfer 0.1 mL of the 5.0 EU/mL endotoxin solution into 0.9 mL of LAL Reagent Water in a suitable container and label 0.5 EU/mL. The solution should be vigorously vortexed for approximately 1 minute before proceeding.



3. Transfer 0.1 ml of the 0.5 EU/mL endotoxin solution into 0.9 mL of LAL Reagent Water in a suitable container and label 0.05 EU/mL. The solution should be vigorously vortexed for approximately 1 minute before proceeding.
4. Transfer 0.1 mL of the 0.05 EU/mL endotoxin solution into 0.9 mL of LAL Reagent Water in a suitable container and label 0.005 EU/mL. The solution should be vigorously vortexed for approximately 1 minute before proceeding.

## Testing Procedure

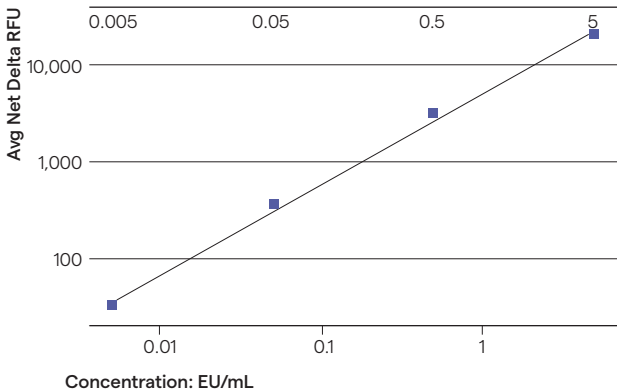
Allow reagents to equilibrate to room temperature prior to use.

### General testing procedure

1. The PyroGene® Assay uses a 96-well plate assay format. Add 100  $\mu\text{L}$  of the blank, endotoxin standards, and samples to appropriate wells of the microplate. Duplicate or triplicate wells are generally used.
2. To spike the samples with endotoxin, choose a spike concentration relative to the possible background endotoxin in the test sample or dilution. For a spike concentration of 0.5 EU/mL, add 10  $\mu\text{L}$  of the 5.0 EU/mL solution to the designated Positive Product Control wells.
3. Pre-incubate the plate in the reader at  $37^{\circ}\text{C} \pm 1^{\circ}\text{C}$  for a minimum of 10 minutes.
4. During the incubation period, prepare the working reagent which consists of fluorogenic substrate, assay buffer and rFC enzyme solution in a 5:4:1 ratio, respectively.
5. Carefully dispense 100  $\mu\text{L}$  of the working reagent to each well.
6. Read fluorescence at time zero.
7. Incubate the reaction for one hour and read the plate at elapsed time one-hour.
8. The difference of time one-hour and time zero readings are corrected with the blank.

- The log net  $\Delta$ RFU is then plotted against log endotoxin concentration in a linear regression curve – an example is shown below.
- Calculate endotoxin concentration of samples according to the standard curve.

### Endotoxin standard curve in the PyroGene® Assay



## When Using PyroWave® XM and WinKQCL® Software

The PyroWave® XM Fluorescence Reader and WinKQCL® Software can be used in this assay.

1. Turn on the reader and launch the software. In the software, create a template for the PyroGene® rFC Assay. Several types of assays may be programmed. Consult the Software Manual or Help for more information. Save the template.
2. From the list of the templates, an assay is initiated by selecting Run. Follow on screen prompts to begin the test. Consult Software Manual or Help for more details.
3. Place the microplate (loaded with blank, standards and samples) in the reader and pre-incubate the plate for a minimum of 10 minutes at  $37^{\circ}\text{C} \pm 1^{\circ}\text{C}$ .
4. During the 10 minute incubation, prepare working reagent by mixing the fluorogenic substrate, the assay buffer and the rFC enzyme solution in a 5:4:1 ratio, respectively. To ensure enough working reagent is prepared for the assay and excess is minimal, determine the number of wells requiring working reagent, add 4 additional wells, and calculate the appropriate amount for each component to make the working reagent. Allow reagents to equilibrate to room temperature before mixing. The order of mixing should remain consistent. Add enzyme last to the buffered substrate. Mix gently but thoroughly. Do not vortex. Refer to the table on the following page. Note: Once prepared, the working reagent cannot be stored.

## Volume ( $\mu\text{L}$ ) of Reagent Required per Total Number of Wells

To assure slight excess, assume volume prepared is sufficient for an additional 4-wells

Total Number of Wells	Fluorogenic Substrate	rFC Assay Buffer	rFC Enzyme Solution	Total Volume
6	500	400	100	1000
12	800	640	160	1,600
24	1,400	1,120	280	2,800
36	2,000	1,600	400	4,000
42	2,300	1,840	460	4,600
48	2,600	2,080	520	5,200
54	2,900	2,320	580	5,800
60	3,200	2,560	640	6,400
66	3,500	2,800	700	7,000
72	3,800	3,040	760	7,600
78	4,100	3,280	820	8,200
84	4,400	3,520	880	8,800
90	4,700	3,760	940	9,400
96	5,000	4,000	1,000	10,000

- When prompted by the software at the end of the incubation period, dispense 100  $\mu\text{L}$  of working reagent into the appropriate wells using an eight channel pipettor.
- Immediately follow screen prompt. The software will initiate read one (time zero reading) followed by read two an hour later.
- After the assay is completed, the data is automatically saved to the hard drive.

## Types of PyroGene® Assays

The incubating fluorescence microplate reader and WinKQCL® Software are an integral part of the PyroGene® Assay. It is important to become familiar with the operation of the incubating microplate reader and the features of the WinKQCL® Software. Please refer to the incubating microplate reader and WinKQCL® Software Manuals or Help for more detailed information.

There are four (4) basic types of PyroGene® Assays. Each type is designed to perform a different aspect of endotoxin testing.

### 1. Routine

A Routine assay calculates the concentration of endotoxin in unknowns by comparison to the performance of a series of endotoxin standards.

As part of a Routine assay, the user has the option to include a Positive Product Control (PPC) as a monitor for product inhibition or enhancement. A PPC is a sample of product to which a known amount of endotoxin spike has been added. The WinKQCL® Software automatically calculates the amount of endotoxin recovered in the PPC, allowing for a comparison to the known amount of endotoxin spike.

### 2. Inhibition/Enhancement

The PyroGene® Reaction is enzyme mediated and, as such, has an optimal pH range and specific salt and divalent cation requirements. Occasionally, test samples may

alter these optimal conditions to an extent that the rFC is rendered insensitive to endotoxin. Negative results with samples which inhibit the PyroGene® Test do not necessarily indicate the absence of endotoxin.

An Inhibition/Enhancement assay is designed to determine what level of product dilution overcomes inhibition or enhancement. Each product dilution must be accompanied by a Positive Product Control (PPC). The WinKQCL® Software calculates the amount of endotoxin recovered in the PPC for comparison to the known amount of endotoxin spike. In this manner, it can be determined which product dilutions are non-inhibitory.

### **3. RSE/CSE**

An RSE/CSE assay is designed to determine the potency of a Control Standard Endotoxin (CSE) in terms of the concentration units of the Reference Standard Endotoxin (RSE).

The assay requires a single series of RSE dilutions and one or more sets of dilutions of the CSE. Depending on the concentration units of the CSE, the WinKQCL® Software automatically computes mean potency values in terms of EU/mL or EU/ng. The user also has the option to enter units other than EU or ng.

### **4. Initial Qualification**

An Initial Qualification assay is designed according to the requirements of the USP Chapter <85> Bacterial Endotoxins Test 12. This assay is required as part of the validation of the LAL assay and is also to be performed with each new

kit lot. PyroGene® Testing evolved from the LAL test and Lonza suggests a similar approach be utilized.

The Initial Qualification assay performs a log/log linear correlation of the individual net  $\Delta$ RFU values for each replicate of each endotoxin standard. The other assays use the average net  $\Delta$ RFU of all the replicates of each standard.

The Initial Qualification assay does not provide for the inclusion of any samples.

## Performance Characteristics

### Linearity\*

The linearity of the standard curve within the concentration range used to determine endotoxin values should be verified. Assay no less than 3 endotoxin standards, spanning the desired concentration range, and a LAL Reagent Water blank at least in triplicate. Additional standards should be included to bracket each log interval over the range of the standard curve. The value of the correlation coefficient (r) of the calculated standard curve should be  $\geq 0.980$ .

\*See additional data considerations under Other Technical Issues section.



## Product Inhibition

Product inhibition occurs when substances in the test sample interfere with the enzyme reaction. Inhibition results in a lower final  $\Delta$ RFU in a sample reading, indicating lower levels of endotoxin than what may actually be present in the test sample. The lack of product inhibition should be determined for each specific sample, either undilute or diluted to an appropriate dilution.

To verify the lack of product inhibition, an aliquot of a test sample (or a dilution of a test sample) is spiked with a known amount of endotoxin. It is recommended that the endotoxin spike be chosen relative to the possible background endotoxin in the test sample or dilution. The spiked solution is assayed along with the unspiked samples and their respective endotoxin concentrations are determined. The differences between these two calculated endotoxin values should equal the known concentration of the spike within the range of 50–200%<sup>12</sup>.

A spiked aliquot of the test sample (or dilution) may be prepared as in one of the following examples:

### **Tube Method:**

Transfer 0.5 mL of the 5.0 EU/mL solution into 4.50 mL of test sample (or dilution). This solution contains an endotoxin concentration of 0.5 EU/mL in test sample (or dilution). This sample should be vigorously vortexed for one minute prior to use.

## Plate Method #1:

Transfer 10  $\mu\text{L}$  of the 5.0 EU/mL solution into each of the PPC wells in the 96-well plate, as directed by the assay template. To these wells, add 100  $\mu\text{L}$  of test sample (or dilution). Each well will now contain a 0.5 EU/mL solution. Mix gently.

## Plate Method #2:

Place 100  $\mu\text{L}$  of test sample (or dilution) into the PPC wells in the 96-well plate, as directed by the assay template. To these wells, add 10  $\mu\text{L}$  of the 5.0 EU/mL solution. Each well will now contain a 0.5 EU/mL solution. Mix gently.

If the test sample (or dilution) is found to be inhibitory to the enzyme reaction, the sample may require further dilution until the inhibition is overcome.

### Determination of a Non-Inhibitory Dilution with 0.5 EU/mL Endotoxin Spike

Sample Dilution Factor (DF)	Raw Endotoxin (EU/mL)			Spike Recovery	Sample Inhibition	Endotoxin (Raw x DF)
	Unspiked	Spiked	Difference			
1/10	0.075	0.1	0.025	5%	Inhibitory	N/A
1/20	0.044	0.264	0.22	44%	Inhibitory	N/A
24	1,400	1,120	280	2,800	Non-Inhibitory	1.0 EU/mL

Initially, Lonza recommends screening for product inhibition by testing ten-fold dilutions of the test sample. Once the approximate non-inhibitory dilution is determined, the exact dilution can be found by testing two-fold dilutions around this dilution.

## Other Technical Issues

### Improving Spike Recovery

By using a different data treatment procedure (for example, a polynomial curve fit using PowerCurve™), calculation of the recovery of the positive product control can be improved.

### Overcoming Inhibition with a Dispersing Agent

PYROSPERSE® Dispersing Agent, Catalog No. N188, is not compatible with the PyroGene® Assay.

## References

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## Patent Information

Various components of the PyroGene® Assay are protected under the following patents:

Country	Patent/Pub No.	Country	Patent/Pub No.
US	5712144	DK	1409984
US	5716836	EP	1409984
US	5858706	ES	1409984
US	5985590	FI	1409984
US	5998389	FR	1409984
US	6849426	GB	1409984
UK	1015609	GR	1409984
SG	71946	IE	1409984
IT	1015609	IT	1409984
DE	1015609	JP	1409984
FR	1015609	LU	1409984
AU	2002330860	MX	254052
BE	1409984	NL	1409984
BR	210681	PT	1409984
CH	1409984	SE	1409984
CN	1529711	AT	1409984
CY	1409984	US	6849426
DE	1409984		

# Notes

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