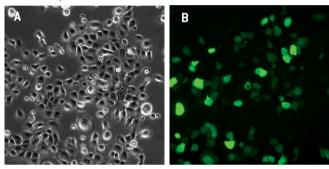


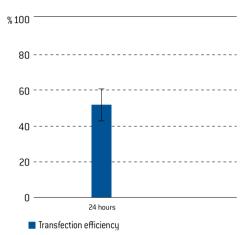
# Amaxa™ Nucleofector™ Protocol for Normal Human Bronchial Epithelial Cells (NHBE)

Validated to work with Clonetics™ NHBE [Lonza; Cat. No. CC-2540]; adherent epithelial cells



**Example for Nucleofection of NHBE Cells** 

NHBE cells were transfected using program W-001 and a plasmid encoding the maxGFP\* fluorescent protein. 24 hours post Nucleofection the cells were analyzed by light (A) and fluorescence microscopy (B).



Average transfection efficiency of NHBE cells 24 hours post Nucleofection. Cells were transfected using Nucleofector™ Program W-001 and 2 µg of pmaxGFP™ Vector. Cell viability is usually around 50%.

# **Product Description**

#### Recommended Kit(s) - Basic Nucleofector™ Kit for Primary Mammalian Epithelial Cells

Cat No.	VPI-1005
Size [reaction]	25
Nucleofector™ Solution	2.25 mL (2.05 mL + 10% overfill)
Supplement	0.5 mL (0.45 mL + 10% overfill)
pmaxGFP™ Vector (0.5 μg/μL in 10 mM Tris pH 8.0)	30 µg
Certified Cuvette	25
Pastic Pipetts	25

#### Storage and Stability

Store Nucleofector<sup>™</sup> Solution, Supplement and pmaxGFP<sup>™</sup> Vector at  $4^{\circ}$ C. For long-term storage, pmaxGFP<sup>™</sup> Vector is ideally stored at -20°C. The expiration date is printed on the solution box. Once the Nucleofector<sup>™</sup> Supplement is added to the Nucleofector<sup>™</sup> Solution it is stable for three months at  $4^{\circ}$ C

#### Note

4D-Nucleofector™ Solutions can only be used with conductive polymer Nucleocuvette™ Vessels, i.e. in the 4D-Nucleofector™ and the 96-well Shuttle™ System. They are not compatible with the Nucleofector™ II/2b Device.

# Required Material

#### Note

Please make sure that the entire supplement is added to the Nucleofector Solution. The ratio of Nucleofector Solution to supplement is 4.5: 1. For a single reaction use 82  $\mu$ L of Nucleofector Solution plus 18  $\mu$ L of supplement to make 100  $\mu$ L of total reaction volume..

- Nucleofector™ Device
- Supplemented Nucleofector™ Solution at room temperature
- Supplied certified cuvettes
- Supplied plastic pipettes
- Supplied pmaxGFP™ Vector
- Substrate of interest, highly purified, preferably by using endotoxin free kits; A260: A280 ratio should be at least 1.8 6-well culture dish or culture system of your choice
- For trypsinization: ReagentPack™ Subculture Reagent Kit containing trypsin/EDTA, HEPES Buffered Saline Solution (HBSS) and Trypsin Neutralizing Solution (TNS) [Lonza, Cat. No. CC-5034]
- Culture medium: BEGM™ BulletKit™ [Lonza; Cat. No. CC-3170])
- Prewarm appropriate volume of culture media at 37°C (1.5 mL per reaction)
- Appropriate number of cells (4–5 x 10<sup>5</sup> cells per sample; minimal cell number: 4 x 10<sup>5</sup> cells; a lower cell number may lead to a major increase in cell mortality

### 1. Pre Nucleofection

#### Cell culture recommendations

- 1.1 Seeding conditions: at least 3.5 x 10<sup>3</sup> cells/cm<sup>2</sup>
- 1.2 Replace media 1 day after splitting, then every 2 days
- 1.3 Cells should be passaged every 3 4 days
- 1.4 For Nucleofection cells should be preferably passaged 2 days before
- 1.5 Do not use cells after passage number 8 as this may result in substantially lower gene transfer efficiency and viability
- 1.6 Optimal confluency before Nucleofection: 60 80%

#### **Trypsinization**

- 1.7 Remove media from the cultured cells and wash cells once with HBSS; use at least same volume of HBSS as culture media
- 1.8 Cells are very sensitive to trypsin treatment. For harvesting, incubate the cells 5 6 minutes at 37°C with recommended volume of indicated trypsinization reagent (please see required material); if cells are incubated >7 10 minutes cells may start to clump
- 1.9 Neutralize trypsinization reaction with TNS once the majority of the cells (>90%) have been detached

## 2. Nucleofection

#### One Nucleofection Sample contains

5 x 105 cells

 $1-5~\mu g$  plasmid DNA (in  $1-5~\mu L$   $H_2O$  or TE) or 2  $\mu g$  pmaxGFP" Vector or 30-300~nM siRNA (3 -30~pmol/sample)

100 µL Nucleofector™ Solution

- 2.1 Please make sure that the entire supplement is added to the Nucleofector™ Solution
- 2.2 Prepare 6-well plates by filling appropriate number of wells with 1.5 mL of supplemented culture media and pre-incubate/equilibrate plates in a humidified 37°C/5% CO<sub>2</sub> incubator
- 2.3 Harvest the cells by trypsinization (please see 1.9–1.11)
- 2.4 Count an aliquot of the trypsinized cells and determine cell density
- 2.5 Centrifuge the required number of cells (5 x 105 cells per sample) at 220xg for 5 minutes at room temperature
- 2.6 Resuspend the cell pellet carefully in 100 µL room temperature Nucleofector™ Solution per sample
- 2.7 Combine 100  $\mu$ L of cell suspension with 1 5  $\mu$ g DNA, 2  $\mu$ g pmaxGFP $^{\text{TM}}$  Vector or 30 nM 300 nM siRNA (3 30 pmol/sample) or other substrates
- 2.8 Transfer cell/DNA suspension into certified cuvette; sample must cover the bottom of the cuvette without air bubbles. Close the cuvette with the cap
- 2.9 Select appropriate Nucleofector™ Program W-001 (W-01 for Nucleofector™ | Device)
- 2.10 Insert the cuvette with cell/DNA suspension into the Nucleofector™ Cuvette Holder and apply the selected program
- 2.11 Take the cuvette out of the holder once the program is finished
- 2.12 Add  $\sim 500~\mu L$  of the pre-equilibrated culture media to the cuvette and gently transfer the sample immediately into the 6-well plate (final volume 1.5 mL media per well/sample). Use the supplied pipettes and avoid repeated aspiration of the sample

## 3. Post Nucleofection

3.1 Incubate the cells in a humidified  $37^{\circ}\text{C}/5\%$  CO<sub>2</sub> incubator until analysis. Gene expression or down regulation, respectively, is often detectable after only 4-8 hours

# Additional Information

For an up-to-date list of all Nucleofector™ References, please refer to: www.lonza.com/nucleofection-citations

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