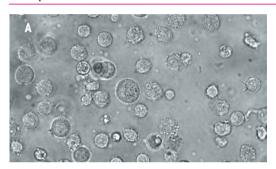


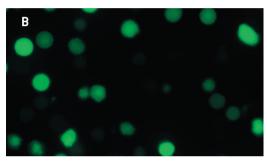
Amaxa® Cell Line Nucleofector® Kit L

For L428 [DSMZ Cat. No ACC-197, cryopreserved]

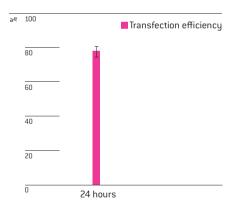
Human Hodgkin Lymphoma; polymorph cells; some cells have spikes

Example for Nucleofection® of L428 cells





L428 cells [DSMZ Cat. No ACC-197] were transfected with the Nucleofector® Kit L, Program X-001 and 2 μ g of pmaxGFP® Vector. Cells were analyzed 24 hours post Nucleofection® using light (A) and fluorescence microscopy (B).



Average transfection efficiency of L428 cells. L428 cells [DSMZ Cat. No ACC-197] were transfected with program X-001 and 2 μg of pmaxGFP® Vector. Cells were analyzed 24 hours post Nucleofection® by flow cytometry. Cell Viability (compared to non-transfected control) is around 73% 24 hours post Nucleofection®.

Product Description

Cat. No.		VCA-1005
Size (reactions)		25
Cell Line Nucleofector® Solution L		2.25 ml (2.05 ml + 10% overfill)
Supplement		0.5 ml (0.45 ml + 10% overfill)
pmaxGFP® Vector (0.5 µg/µl in 10 mM Tris pH 8.0)		30 µg
Certified cuvettes		25
Plastic pipettes		25
Storage and stability	Store Nucleofector® Solut	tion Supplement and pmayCEP® Vector at 1°C. For long-term storage

Storage and stability

Store Nucleofector® Solution, Supplement and pmaxGFP® Vector at 4°C. For long-term storage, pmaxGFP® Vector is ideally stored at -20°C. The expiration date is printed on the solution box. Once the Nucleofector® Supplement is added to the Nucleofector® Solution it is stable for three months at 4°C.

Optimized Protocol for L428 Cell Line [DSMZ]

Required Material

Note

Please make sure that the entire supplement is added to the Nucleofector® Solution. The ratio of Nucleofector® Solution to supplement is 4.5:1. For a single reaction use 82 μ l of Nucleofector® Solution plus 18 μ l of supplement to make 100 μ l of total reaction volume.

- Nucleofector® Device, (Software requirements: version V2.3 or higher for Nucleofector® I Device; version S3.4 or higher for Nucleofector® II Device)
- Supplemented Nucleofector® Solution at room temperature
- Supplied certified cuvettes
- Supplied plastic pipettes
- Supplied pmaxGFP® Vector
- Substrate of interest, highly purified, preferably by using endotoxin-free kits; A260: A280 ratio should be at least 1.8
- 12-well culture dish or culture system of your choice
- Culture medium: RPMI 1640 supplemented with 2 mM L-glutamine; fetal bovine serum, 10%
- Prewarm appropriate volume of culture medium to 37°C (1 ml per sample)
- Appropriate number of cells (1 x 10^6 cells per sample; lower or higher cell numbers may influence transfection results)

1. Pre Nucleofection®

Cell culture recommendations

- 1.1 Replace media every 2 3 days
- 1.2 Passage cells every 2 3 days. A subculturing ratio of 1:2 to 1:3 is recommended
- 1.3 Seed out $2.5 3.5 \times 10^5 \text{ cells/ml}$
- 1.4 Subculture 2-3 days before Nucleofection® with a ratio of 1:2-1:3

Optimized Protocol for L428 Cell Line [DSMZ]

2. Nucleofection®

One Nucleofection® Sample contains

 1×10^6 cells

 $2 \mu g$ plasmid DNA (in $1-5 \mu l$ H₂0 or TE) or $2 \mu g$ pmaxGFP® Vector or 30-300nM siRNA $(3-30 \mu l)$ pmol/sample)

100 µl Cell Line Nucleofector® Solution L

- 2.1 Please make sure that the entire supplement is added to the Nucleofector® Solution
- 2.2 Prepare 12-well plates by filling appropriate number of wells with 1 ml of supplemented culture media and pre-incubate/equilibrate plates in a humidified 37°C/5% CO₂ incubator
- 2.3 Count an aliquot of the cells and determine cell density
- 2.4 Centrifuge the required number of cells (1 x 10^6 cells per sample) at 90xg for 10 minutes at room temperature. Remove supernatant completely
- 2.5 Resuspend the cell pellet carefully in 100 µl room-temperature Nucleofector® Solution per sample

Note Avoid leaving the cells in Nucleofector® Solution for extended periods of time (longer than 15 minutes), as this may reduce cell viability and gene transfer efficiency.

- 2.6 Combine 100 μ l of cell suspension with **2 \mug** DNA, 2 μ g pmaxGFP® Vector or **30 nM 300 nM** siRNA (3 30 pmol/sample) or other substrates
- 2.7 Transfer cell/DNA suspension into certified cuvette (sample must cover the bottom of the cuvette without air bubbles). Close the cuvette with the cap
- 2.8 Select the appropriate Nucleofector® Program X-001 (X-01 for Nucleofector® | Device)
- 2.9 Insert the cuvette with cell/DNA suspension into the Nucleofector® Cuvette Holder and apply the selected program by pressing the X-button
- 2.10 Take the cuvette out of the holder once the program is finished
- 2.11 Incubate the sample in the cuvette for 10 minutes at room temperature
- 2.12 Add \sim 500 μ l of the pre-equilibrated culture medium to the cuvette and gently transfer the sample into the prepared 12-well plate (final volume 1.5 ml media per well). Use the supplied pipettes and avoid repeated aspiration of the sample

3. Post Nucleofection®

3.1 Incubate the cells in humidified 37° C/5% CO₂ incubator until analysis. Gene expression or down regulation, respectively, is often detectable after only 4-8 hours

2009-05

Optimized Protocol for L428 Cell Line [DSMZ]

Additional Information

For an up-to-date list of all Nucleofector® References, please refer to: www.lonza.com/nucleofection-citations

For more technical assistance, contact our Scientific Support Team:

USA/Canada Europe and Rest of World

Phone: 800 521 0390 (toll-free) Phone: +49 221 99199 400

Fax: 301 845 8338 Fax: +49 221 99199 499

Lonza Cologne AG 50829 Cologne, Germany

 $Please \ note that \ the \ Amaxa^{@}\ Nucleofector^{@}\ Technology\ is\ not\ intended\ to\ be\ used\ for\ diagnostic\ purposes\ or\ for\ testing\ or\ treatment\ in\ humans.$

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